

PATHOZYME[®] PROSTATE SPECIFIC ANTIGEN Ref OD327

Enzyme-Immunoassays (EIA) for the detection of PSA in human serum

Store at 2°C to 8°C. DO NOT FREEZE

For in-vitro use only.

INTRODUCTION

Human Prostate Specific Antigen (PSA) is a serine protease, a single chain glycoprotein with a molecular weight of approximately 34,000 daltons containing 7% carbohydrate by weight. PSA is immunologically specific for prostatic tissue. It is also present in normal, benign, hyperplastic and malignant prostatic tissue, in metastatic prostatic carcinoma and also in prostatic fluid and seminal plasma. PSA is not present in any other normal tissue obtained from men, nor is it produced by cancer of the breast, lung, colon, rectum, stomach, pancreas, or thyroid. It is functionally and immunologically different from prostatic acid phosphatase (PAP).

Elevated serum PSA concentrations have been reported in patients with prostate cancer, benign prostatic hypertrophy or inflammatory conditions of other adjacent genito-urinary tissues. Elevated PSA concentrations have not been found in apparently healthy men, men with non-prostatic carcinoma, apparently healthy women or women with cancer. Reports have suggested that serum PSA is one of the most useful markers in oncology. It may serve as an accurate marker for assessing response to treatment in patients with prostatic cancer. Therefore, measurement of serum PSA concentrations can be an important tool in monitoring patients with prostatic cancer and in determining the potential and actual effectiveness of surgery or other therapies. Recent studies also indicate that PSA measurements can enhance early prostate cancer detection when combined with digital rectal examination (DRE).

INTENDED USE

PATHOZYME PSA is an Enzyme-Immunoassay (EIA) for the quantitative determination of Prostate Specific Antigen (PSA) in human serum.

For professional use only.

PRINCIPLE OF THE TEST

Specific, goat anti-PSA antibodies are coated on to microtitration wells. Test sera are applied and incubated with Assay Buffer. Then monoclonal anti-PSA labelled with Horseradish Peroxidase enzyme (Conjugate) is added. Unbound material is washed away. If human PSA is present in the sample, it will combine with the antibody on the well and the enzyme Conjugate, resulting in the PSA molecules being 'sandwiched' between the solid phase and the enzyme linked antibodies. After incubation, the wells are washed with water to remove unbound labelled antibodies. On addition of the Substrate (TMB), a colour will develop only in those wells in which enzyme is present, indicating the presence of PSA. The enzyme reaction is stopped by the addition of Dilute Hydrochloric Acid and the absorbance is then measured at 450nm. The concentration of PSA is directly proportional to the colour intensity of the test sample. This test has been calibrated against in house standards. There is no International standard for this test.

Ref
OD327

CONTENTS



Microtitre Plate	12 x 8 wells x 1
Breakable wells coated with specific goat anti PSA antibodies contained in a resealable foil bag with a desiccant.	
Cal A 0 ng / ml	1 ml
Reference Standard: Human serum free of PSA. Ready to use. (Colourless)	
Cal B 2 ng / ml	1ml
Reference Standard: PSA diluted in human serum. Ready to use. (Colourless)	
Cal C 4 ng / ml	1ml
Reference Standard: PSA diluted in human serum. Ready to use. (Colourless)	
Cal D 15 ng / ml	1ml
Reference Standard: PSA diluted in human serum. Ready to use. (Colourless)	
Cal E 60 ng / ml	1ml
Reference Standard: PSA diluted in human serum. Ready to use. (Colourless)	
Cal F 120 ng / ml	1ml
Reference Standard: PSA diluted in human serum. Ready to use. (Colourless)	
Conj	11 ml
Anti-PSA HRP Conjugate: Monoclonal anti-PSA conjugated to HRP. Ready to use (Red)	
Buf AS	7 ml
Phosphate based buffer containing stabilising proteins. Ready to use. (Green)	
Subs TMB	11 ml
Substrate Solution: 3,3', 5,5' Tetramethyl Benzidine in a citrate buffer. Ready to use. (Colourless)	
Soln Stop HCl 1M	11 ml
Stop Solution: Hydrochloric Acid diluted in purified water. Ready to use. (Colourless)	
Instruction leaflet and EIA Data Recording Sheet	1 + 1

MATERIAL REQUIRED BUT NOT PROVIDED

Micropipettes: 100µl, 200µl and 1000µl
 Disposable pipette tips
 Absorbent paper
 Microplate reader fitted with a 450nm filter
 Graph paper
 Thoroughly clean laboratory glassware

PRECAUTIONS

PATHOZYME PSA contains materials of human origin which have been tested and confirmed negative for HCV, HIV I and II antibodies and HbsAg by FDA approved methods at single donor level. Because no test can offer complete assurance that products derived from human source will not transmit infectious agents it is recommended that the reagents within this kit be handled with due care and attention during use and

disposal. All reagents should, however, be treated as potential Biohazards in use and for disposal. Do not ingest.

PATHOZYME PSA Reagents do not contain dangerous substances as defined by current UK Chemicals (Hazardous Information and Packaging for Supply) regulations. All reagents should, however, be treated as potential biohazards in use and disposal. Final disposal must be in accordance with local legislation.

PATHOZYME PSA Stop Solution is dilute Hydrochloric Acid and is therefore corrosive. Handle with care. In case of contact, rinse thoroughly with water.

PATHOZYME PSA contain 1% Proclin™ 300* as a preservative which may be toxic if ingested. In case of contact, rinse thoroughly with running water and seek medical advice. *Proclin™ 300 is a trade mark of ROHM & HAAS Limited.

STORAGE

Reagents must be stored at temperatures between 2°C to 8°C. Expiry date is the last day of the month on the bottle and the kit label. The kit will perform within specification until the stated expiry date as determined from date of product manufacture and stated on kit and components. Do not use reagents after the expiry date.

Exposure of reagents to excessive temperatures should be avoided. Do not expose to direct sunlight. DO NOT FREEZE ANY OF THE REAGENTS (Except Standards for storage) as this will cause irreversible damage.

SPECIMEN COLLECTION AND PREPARATION

Obtain a sample of venous blood from the patient and allow a clot to form and retract. Centrifuge clotted blood sample and collect clear serum. Fresh serum samples are required.

Do not use haemolysed, contaminated or lipaemic serum for testing as this will adversely affect the results.

Serum may be stored at 2°C to 8°C for up to 48 hours prior to testing. If longer storage is required, store at -20°C for up to 1 year. Thawed samples must be mixed prior to testing.

Do not use Sodium Azide as a preservative as this may inhibit the Peroxidase enzyme system.

Do not repeatedly freeze-thaw the specimens as this will cause false results.

REAGENT PREPARATION

All reagents should be brought to room temperature (20°C to 25°C) and mixed gently prior to use. Do not induce foaming.

LIMITATIONS OF USE

The use of samples other than serum has not been validated in this test. There is no reuse protocol for this product. When making an interpretation of the test it is strongly advised to take all clinical data into consideration. Diagnosis should not be made solely on the findings of one clinical assay.

ASSAY PROCEDURE

- Bring all the kit components and the test serum to room temperature (20°C to 25°C) prior to the start of the assay.
- One set of Standards should be run with each batch of test serum. Secure the desired number of coated wells in the holder. Record the position of the standards and the test serum on the EIA Data Recording Sheet provided.
- Unused strips should be resealed in the foil bag containing the desiccant, using the resealing zip-lock before being replaced at 2°C to 8°C.
- Dispense 50µl of Standards and test serum into the assigned wells.
- Dispense 50µl of Assay Buffer into each well.
- Thoroughly mix for 30 seconds. It is very important to have a complete mixing at this stage.
- Incubate the plate for 60 minutes at room temperature (20°C to 25°C).
- At the end of the incubation period, discard the contents of the wells by flicking plate contents into a Biohazard container. Then strike the wells sharply against absorbent paper. Ensure adequate disinfectant is contained in the Biohazard container.
- Hand Washing: Fill the wells with a minimum of 300µl of distilled water per well. Flick plate contents into a Biohazard container. Then strike the wells sharply against absorbent paper. Wash the empty wells 5 times.
- Strike the wells sharply onto absorbent paper or paper towel to remove all residual water droplets.
- Machine Washing: Ensure that 300µl of distilled water is dispensed per well and that an appropriate disinfectant is added to the waste collection bottle. Wash the empty wells 5 times. After washing remove excess fluid by striking the wells sharply onto absorbent paper or paper towel to remove all residual water droplets.
- Dispense 100µl of Anti-PSA HRP Conjugate into each well. Gently mix for 5 seconds.
- Incubate the plate for 60 minutes at room temperature (20°C to 25°C).
- Wash plate as described above.
- Dispense 100µl of Substrate Solution into each well and gently mix for 10 seconds.
- Incubate in the dark for 20 minutes at room temperature (20°C to 25°C).
- Stop the reaction by adding 100µl Stop Solution to each well.
- Gently mix for 30 seconds to ensure that the blue colour changes completely to yellow colour.
- Read the optical density immediately (no later than 10 minutes) using a microplate reader with a 450nm filter.

TROUBLESHOOTING

For use by operatives with at least a minimum of basic laboratory training.

Do not use damaged or contaminated kit components.

Use a separate disposable tip for each sample to prevent cross contamination.

Duplication of all standards and specimens, although not required, is recommended.

Specimens and standards should be run at the same time to keep testing conditions the same.

It is recommended that no more than 32 wells be used for each assay run if manual pipetting is used, since pipetting of all Standards and specimens should be completed within 3 minutes. A full plate of 96 wells may be used if automated pipetting is available.

Replace caps on all reagents immediately after use.

Avoid repeated pipetting from stock reagents as this is likely to cause contamination.

Do not mix reagents or antibody coated strips from different kits. When dispensing, care should be taken not to touch the surface of the well.

Do not allow reagent to run down the sides of the well. Prior to the start of the assay bring all reagents to room temperature (20°C to 25°C). Gently mix all reagents by gentle inversion or swirling. Once an assay has been initiated, the wells should not be allowed to become dry during the assay.

Do not contaminate the Substrate Solution as this will render the whole kit inoperative.

Check the precision and accuracy of the laboratory equipment used during the procedure to ensure reproducible results.

The unused strips should be resealed in the foil bag, containing the desiccant, using the resealing zip-lock before being replaced at 2°C to 8°C.

CALCULATION OF RESULTS

Calculate the mean absorbance value (A450) for each set of Standards and specimens. Construct a standard curve by plotting the mean absorbance from each Standard against its concentration in ng/ml on graph paper. Use the mean absorbance values for each specimen to determine the corresponding concentration of PSA in ng/ml from the standard curve. If levels of controls or users known samples do not give expected results, test results must be considered invalid. If using a software package choose a quadratic regression curve fit.

EXPECTED VALUES AND SENSITIVITY

The graph produced by the calibrators should be Hyperbolic in shape with the OD450 of the calibrators is proportional to their concentration. The OD of Calibrator A should be less than 0.75 and the OD of Calibrator F should be greater than 1.5 for the assay results to be valid. Healthy males are expected to have PSA values below 4 ng/ml. The minimum detectable concentration of PSA by **PATHOZYME PSA** is estimated to be 0.25 ng/ml.

EVALUATION DATA

Calibrated to major competitors and in house standards. The co-efficient of variation of **PATHOZYME PSA** is less than or equal to 10%.

In an evaluation between the Omega Pathozyyme PSA kit and the Abbott AxSym PSA kit for samples with levels between 0.1 and 1410 ng/ml the following data was generated.

Number of Samples	161
Correlation Co-efficient	0.987
Slope	0.958
Intercept	0.04
Omega Mean	47.72 ng/ml
Abbott Mean	45.78 ng/ml

In an evaluation between the Omega Pathozyyme PSA kit and the DSL Active PSA kit for samples with levels between 0.1 and 1320 ng/ml the following data was generated.

Number of Samples	161
Correlation Co-efficient	0.993
Slope	0.951
Intercept	0.61
Omega Mean	47.72 ng/ml
DSL Mean	45.99 ng/ml

In both studies the kits gave good correlation.

REFERENCES

- (1) Hara, M. and Kimura, H. Two prostate specific antigens, gamma-seminoprotein and beta-microseminoprotein. *J. Lab. Clin. Med.* 113:541-548;1989.
- (2) Yuan, J. J., Coplen, D. E., Petros, J. A., Figenshau, R. S., Ratliff, T. L., Smith, D. S. and Catalona, W. J. Effects of rectal examination, prostatic massage, ultra-sonography and needle biopsy on serum prostate specific antigen levels. *J. Urol.* 147:810-814;1992.
- (3) Wang, M. C., Papsidero, L. D., Kuriyama, M., Valenzuela, L. A., Murphy, G. P. and Chu, T. M. Prostatic antigen: a new potential marker for prostatic cancer. *Prostate* 2:89-93;1981.
- (4) Stowell, L. I., Sharman, I. E. and Hamel, K. An Enzyme-Linked Immunosorbent Assay (ELISA) for Prostate-specific antigen. *Forensic Science Intern.* 50:125-138; 1991.
- (5) Frankel, A. E., Rouse, R. V., Wang, M. C., Chu, T. M. and Herzenberg, L. A. Monoclonal Antibodies to a human prostate antigen. *Canc. Res.* 42:3714;1982.
- (6) Benson, M. C., Whang, I. S., Pantuck, A., Ring, K., Kaplan, S. A., Olsson, C. A. and Cooner, W. H. Prostate specific antigen density: a means of distinguishing benign prostatic hypertrophy and prostate cancer. *J. Urol.* 147:815-816;1992.
- (7) Gorman, C. The private pain of prostate cancer. *Time.* 10(5):77-80; 1992.
- (8) Walsh, P. C. Why make an early diagnosis of prostate cancer. *J. Urol.* 147:853-854;1992.
- (9) Labrie, F., Dupont, A., Suburu, R., Cusan, L., Tremblay, M., Gomez, J.-L. and Emond, J. Serum prostate specific antigen as pre-screening test for prostate cancer. *J. Urol.* 147:846-852; 1992.
- (10) McCarthy, R. C., Jakubowski, H. V. and Markowitz, H. Human prostate acid Phosphatase: purification, characterisation, and optimisation of conditions for radioimmunoassay. *Clin. Chem. Acta.* 132:287-293;1983
- (11) Heller, J. E. Prostatic acid phosphatase: Its current clinical status. *J. Urol.* 137:1091-1099;1987.
- (12) Filella, X., Molina, R., Umbert, J. J. B., Bedini, J. L. and Ballesta, A. M. Clinical usefulness of prostate specific antigen. *Tumour Biol.* 11:289-294; 1990.
- (13) Shin, W. J., Gross, K., Mitchell, B., Collins, J., Wierzbinski, B., Magoun, S. and Ryo, U. Y. Prostate adenocarcinoma using Gleason scores correlates with prostate-specific antigen and prostate acid phosphatase measurements. *J. Nat. Med. Assoc.* 84:1049-1050; 1992.
- (14) Wirth, M. P. and Frohmuller, H. G. Prostate specific antigen and prostate acid phosphatase in the detection of early prostate cancer and in the prediction of regional lymph node metastases. *Eur. Urol.* 21:263-268; 1992.
- (15) Campbell, M. L. More cancer found with sensitive PSA assay. *Urol. Times.* 20:10; 1992.
- (16) Brawer, M. K., Chetner, M. P., Beattie, J., Buchner, D. M., Vessella, R. L. and Lange, P. H. Screening for prostatic carcinoma with prostate specific antigen. *J. Urol.* 147:841-845; 1992.
- (17) Benson, M. C., Whang, I. S., Olsson, C. A., McMahon, D. J. and Cooner, W. H. The use of prostate specific antigen density to enhance the predictive value of intermediate levels of serum prostate specific antigen. *J. Urol.* 147:817-821; 1992.
- (18) Oesterling, J. E. and Hanno, P. M. PSA still finding niches in cancer diagnosis. *Urol. Times.* 20:13-18; 1992.
- (19) Babaian, R. J., Fritsche, H. A. and Evans, R. B. Prostate-specific antigen and the prostate gland volume: correlation and clinical application. *J. Clin. Lab. Anal.* 4:135-137; 1990.
- (20) Vessella, R.L.; Noteboom, J, and Lange, P.H. Evaluation of the Abbott IMx Automated Immunoassay of Prostate Specific Antigen. *Clin Chem* 38: 2044-2054;1992.

QUICK REFERENCE TEST PROCEDURE

1. Dispense 50µl of test serum or standards and 50µl of Assay Buffer into each well. Gently mix for 30 seconds.
2. Incubate for 60 minutes at room temperature (20°C to 25°C).
3. Discard well contents and wash five times with distilled water.
4. Dispense 100µl of Anti-PSA HRP Conjugate into each well and gently mix for 5 seconds.
5. Incubate for 60 minutes at room temperature (20°C to 25°C).
6. Discard well contents and wash 5 times with distilled water.
7. Add 100µl Substrate Solution to each well and gently shake for 10 seconds.
8. Incubate in the dark for 20 minutes at room temperature (20°C to 25°C).
9. Add 100µl of Stop Solution to each well and gently shake for 30 seconds.
10. Read the Optical Densities immediately (no later than 10 minutes) using microplate reader with a 450nm filter.

8082 ISSUE 6B Revised December 2011
© Omega Diagnostics Ltd., 2011

